

The SOFAST-HMQC

- theorie and application -

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The logo consists of the letters 'FEMP' in a bold, sans-serif font. The 'F' is white and positioned to the left of the 'E', which is black. The 'M' and 'P' are also black and positioned to the right of the 'E'. The letters are closely spaced and have a slight shadow effect.

Introduction

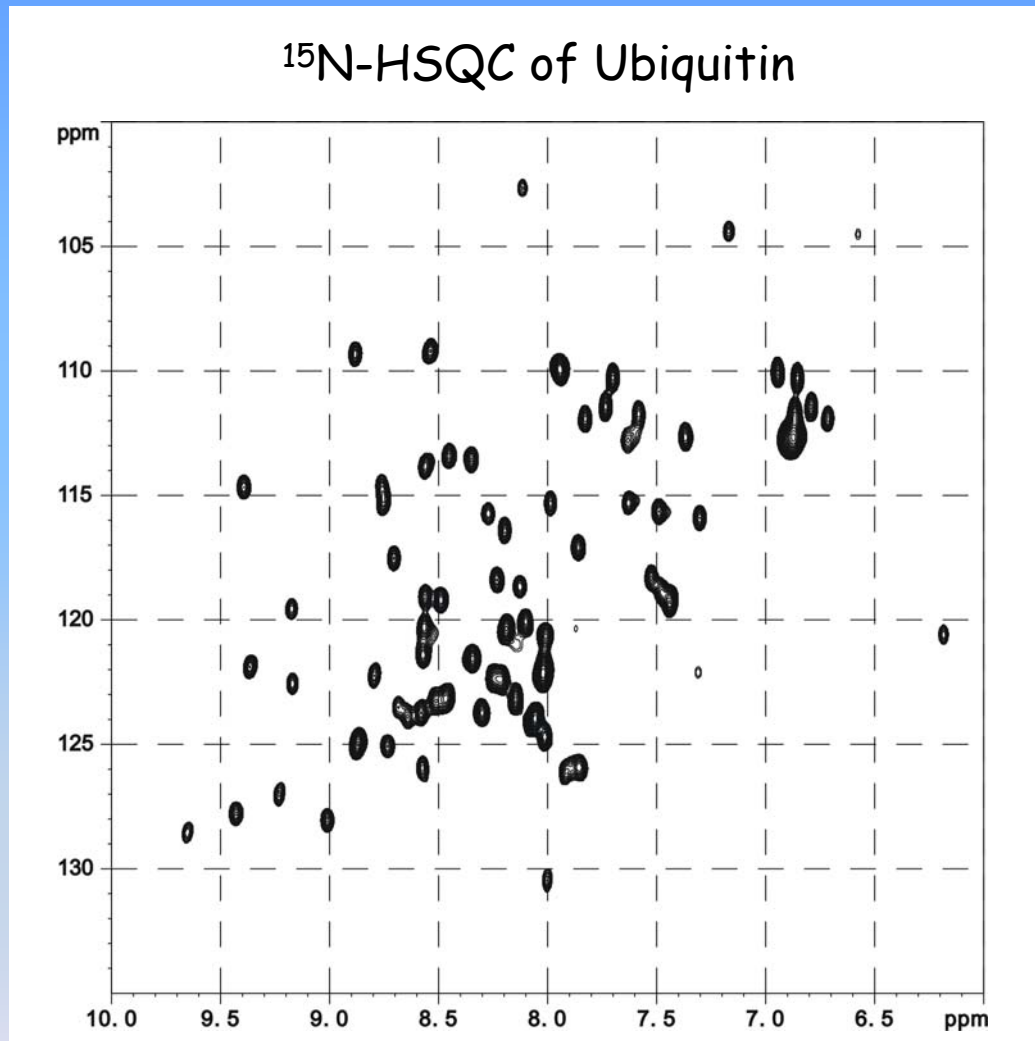
The Ernst angle

Longitudinal Relaxation

SOFAST-HMQC

Ubiquitin

„Nascent chain“



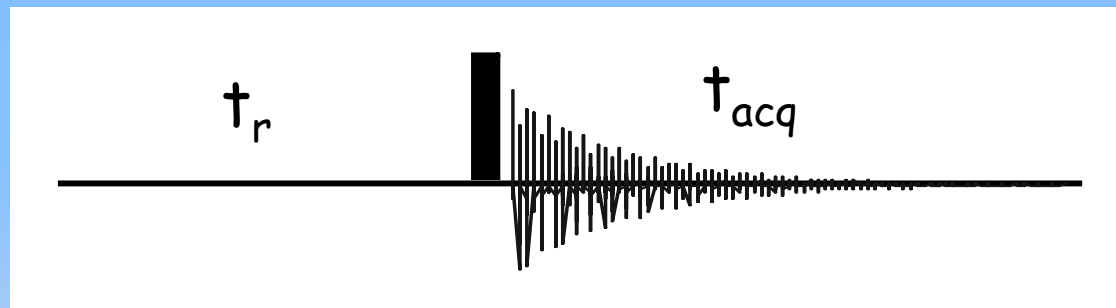
An $^1\text{H}, ^{15}\text{N}$ -
correlation is
of central
importance in
protein NMR

An $^1\text{H}, ^{15}\text{N}$ -correlation

- is a „fingerprint“ of the protein
- is a good test whether the protein is folded or not
- is the seed spectrum for sequence specific assignment
- is used for relaxation measurements
- is used for detection of interactions
- is used for folding experiments
- is a very sensitive experiment (given ^{15}N -labeling)

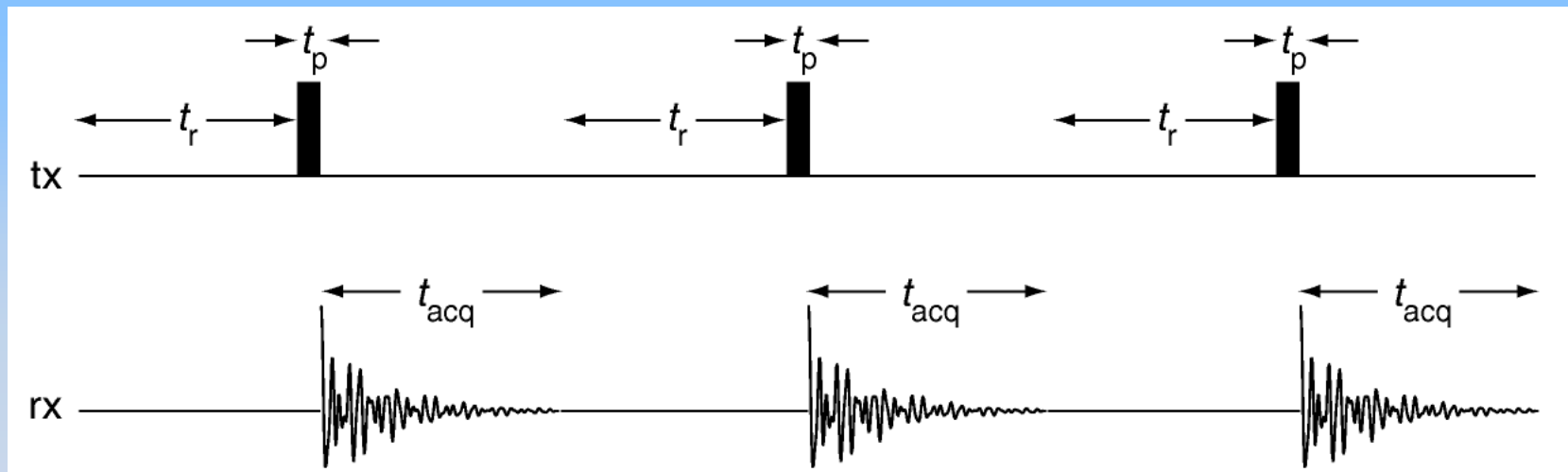
*it would be desirable to be able
to record it fast*

This poses the question how we can achieve a maximum signal-to-noise in a minimum time



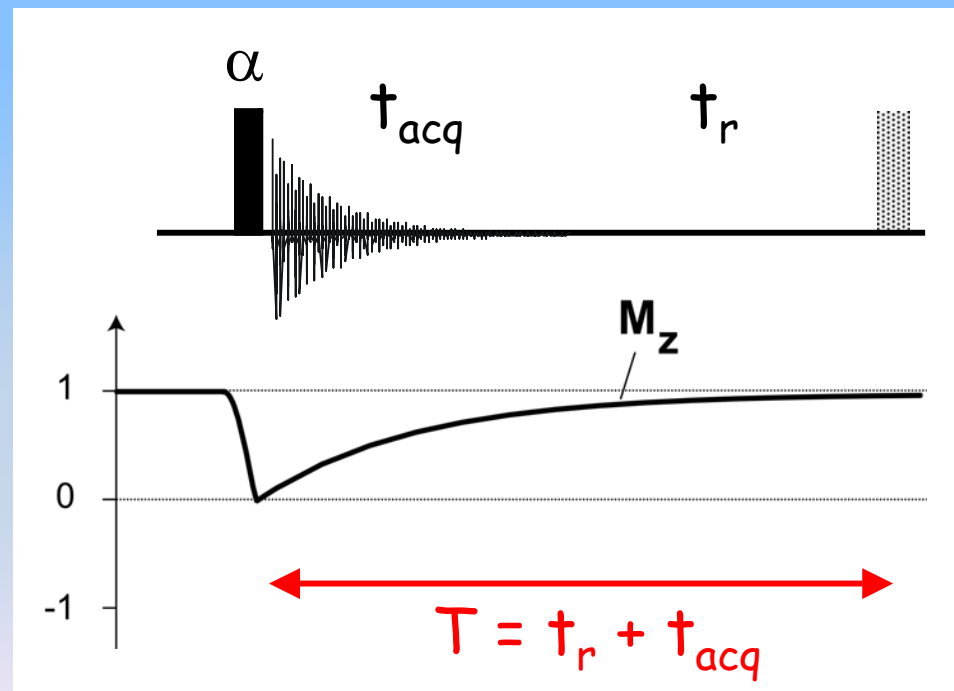
For a simple one-dimensional spectrum this question has been answered by Ernst

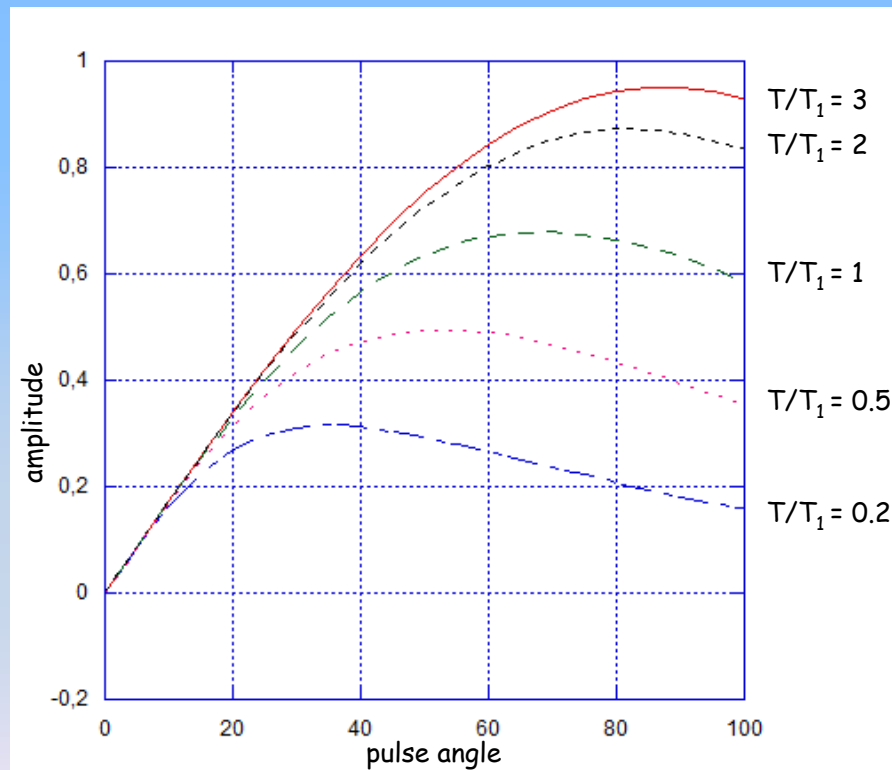
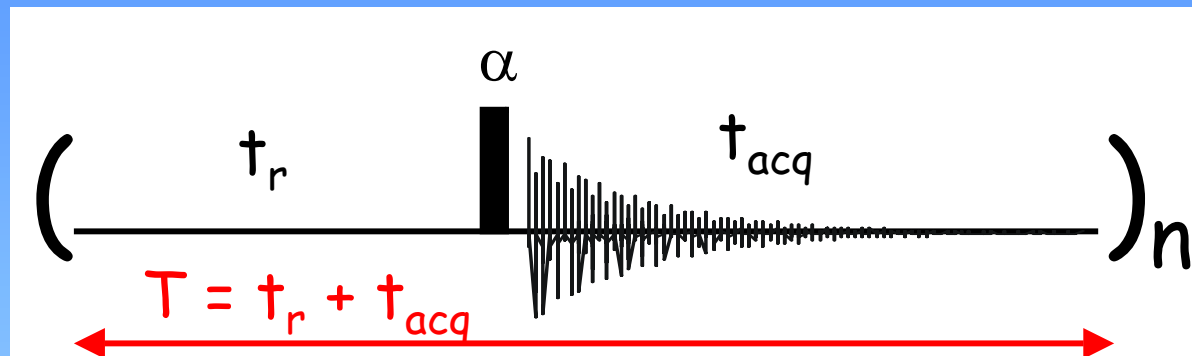
If only one pulse would be applied, the best experiment would use a 90° pulse. With that a maximum of magnetisation would be transferred in the x,y -plane. But because of S/N usually more than one scan is performed



$$T = t_r + t_{acq}$$

After the pulse z-magnetization is recovering with a typical rate, the T_1 -relaxation time. This magnetization is used for the next pulse and the relation between the pulse angle α and the ratio T/T_1 is important



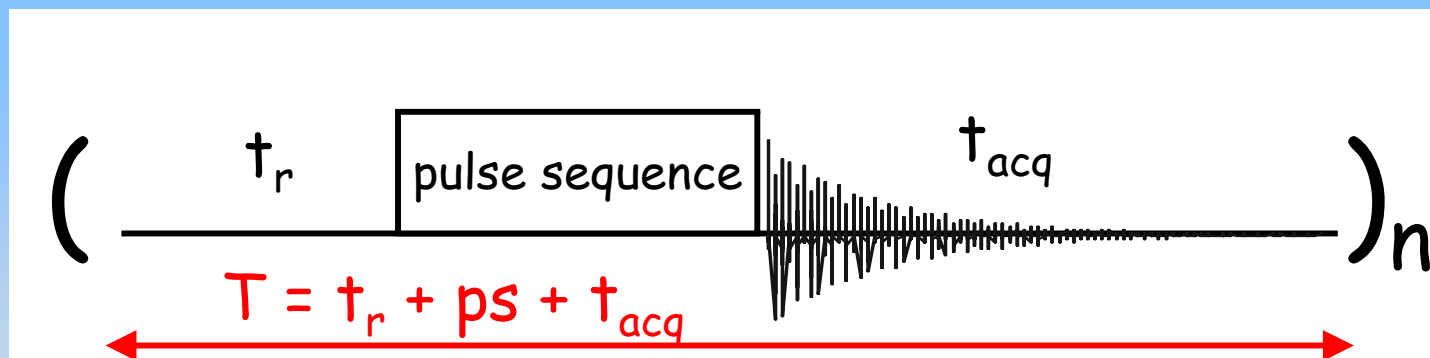


The result is the „Ernst angle“:

$$\cos \alpha = \exp(-T/T_1)$$

that gives best S/N per time (but not necessarily realistic integrals)

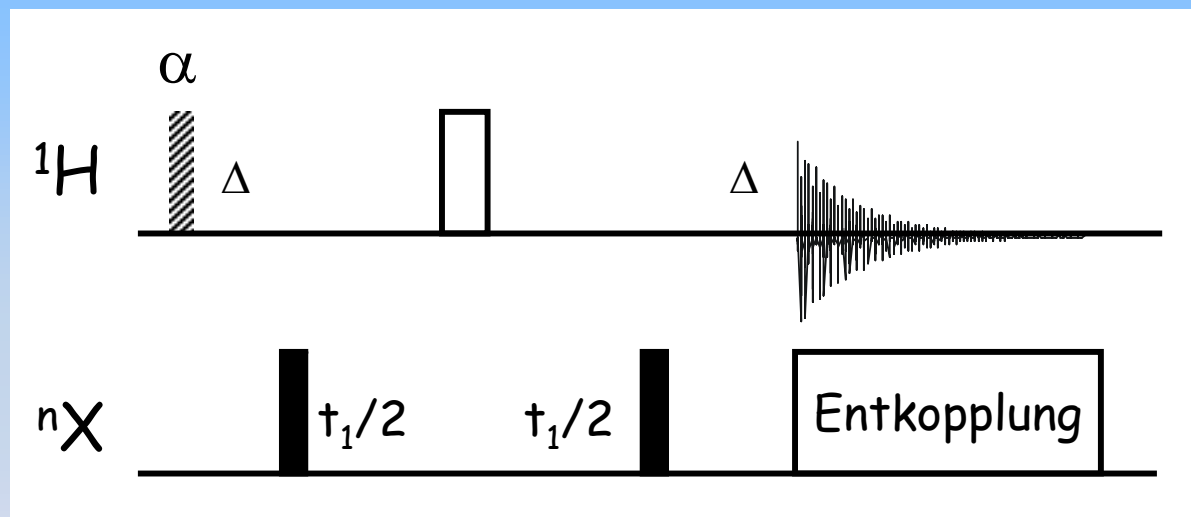
For multidimensional experiments the situation is more complicated since the pulse is now replaced by a complex pulse sequence



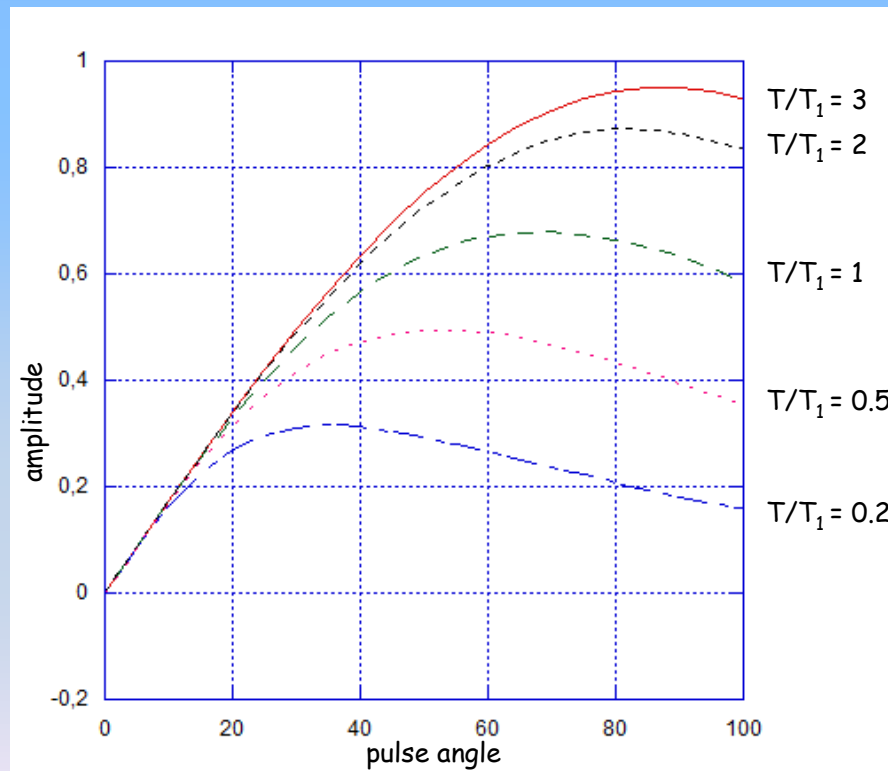
This has been analyzed in detail by

A.Ross, M. Salzmann, H. Senn *J. Biomol. NMR* 10, 389-396 (1997)

They find that for most sequences the 90° pulse is the best option, but for an HMQC they derive an initial pulse of 120° to allow for fast pulsing with 200 msec recovery delay: The Fast-HMQC



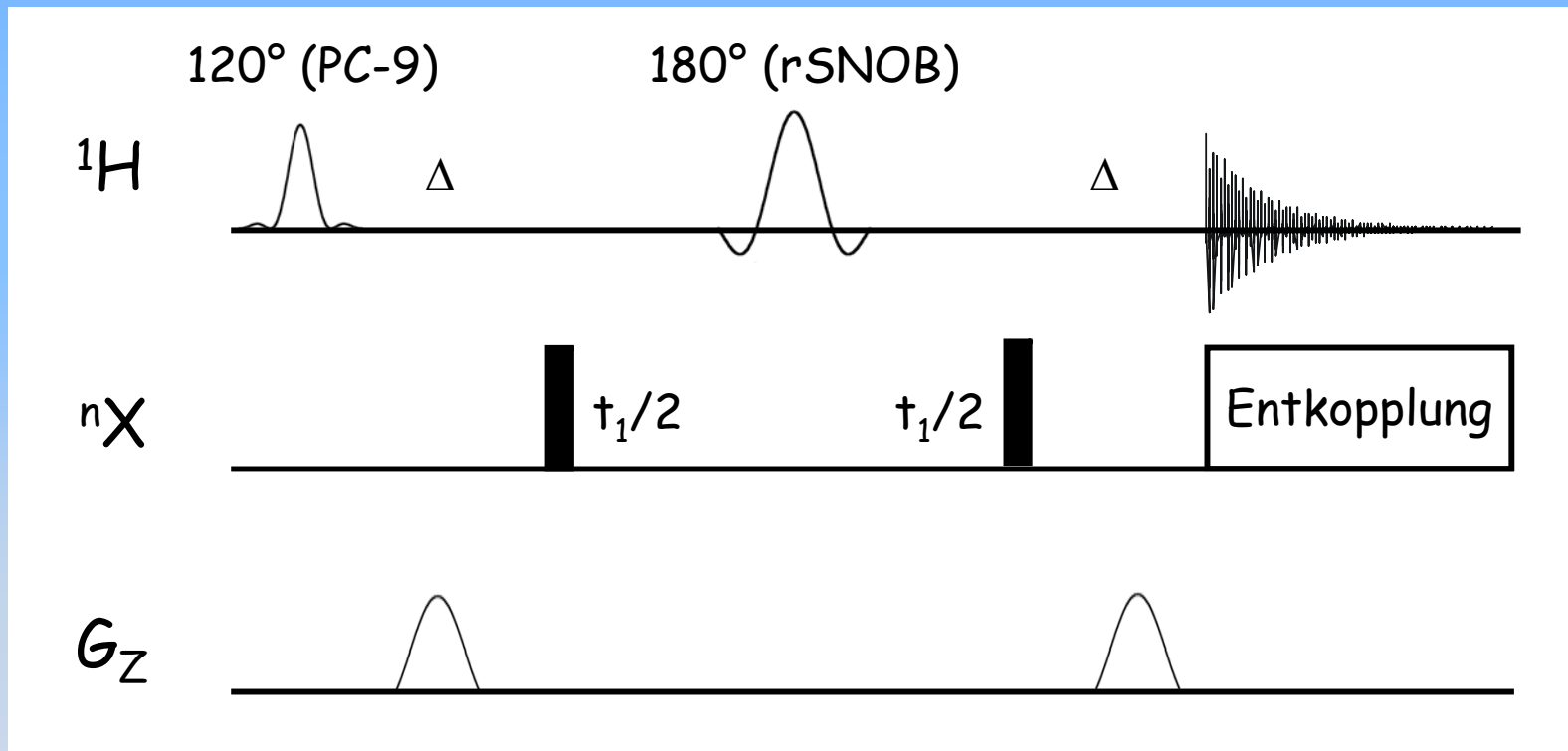
Obviously the value for the relaxation time T_1 has an influence on the repetition rate. For a given pulse angle a shorter T_1 makes a shorter T possible



There are several examples of shortening the T_1 relaxation time, e.g. in the vicinity of paramagnetic centers. This is, however, not generally applicable. But it was shown that an excitation of the amide protons without exciting the other protons in a protein can considerably shorten the relaxation as compared to an unselective excitation

K. Pervushin, B. Vögeli, A. Eletsy *J.Am.Chem.Soc.* **124**, 12898-12902 (2002)

The SOFAST-HMQC takes up both tricks: the Ernst-angle and the reduction of T_1



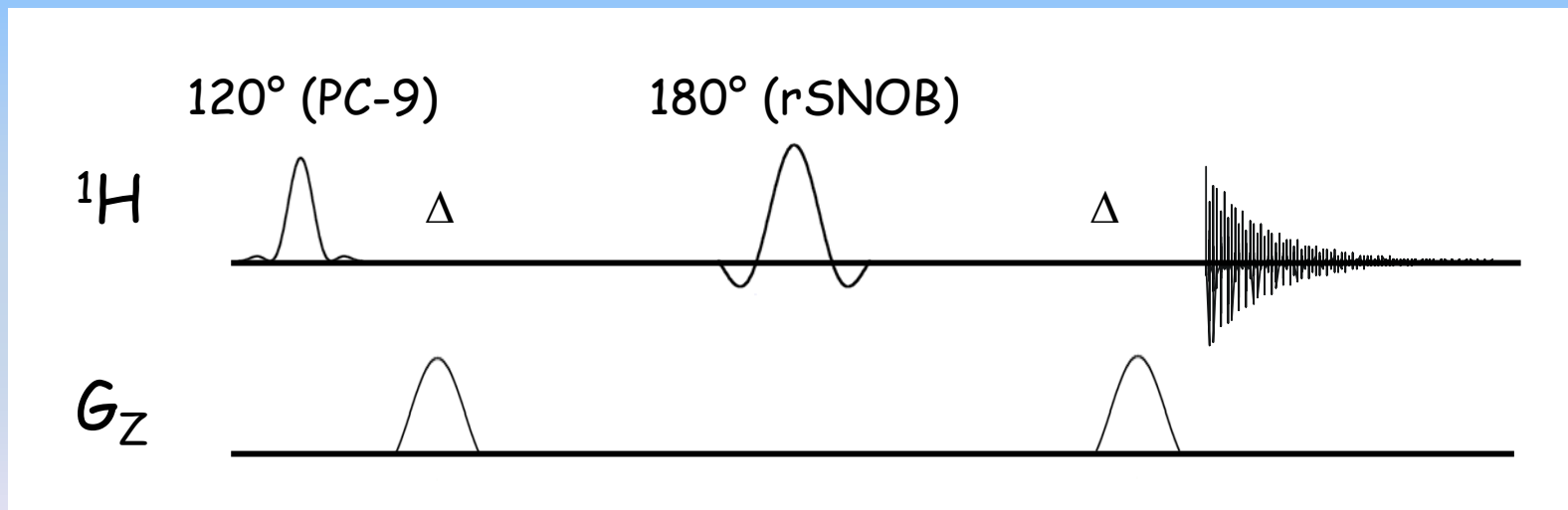
P. Schanda, B. Brutscher *J. Am. Chem. Soc.* **127**, 8014-8015 (2005)
 P. Schanda, E. Kupce, B. Brutscher *J. Biomol. NMR* **33**, 199-211 (2005)

The selective pulses are adjusted so that only the region of amino protons is excited

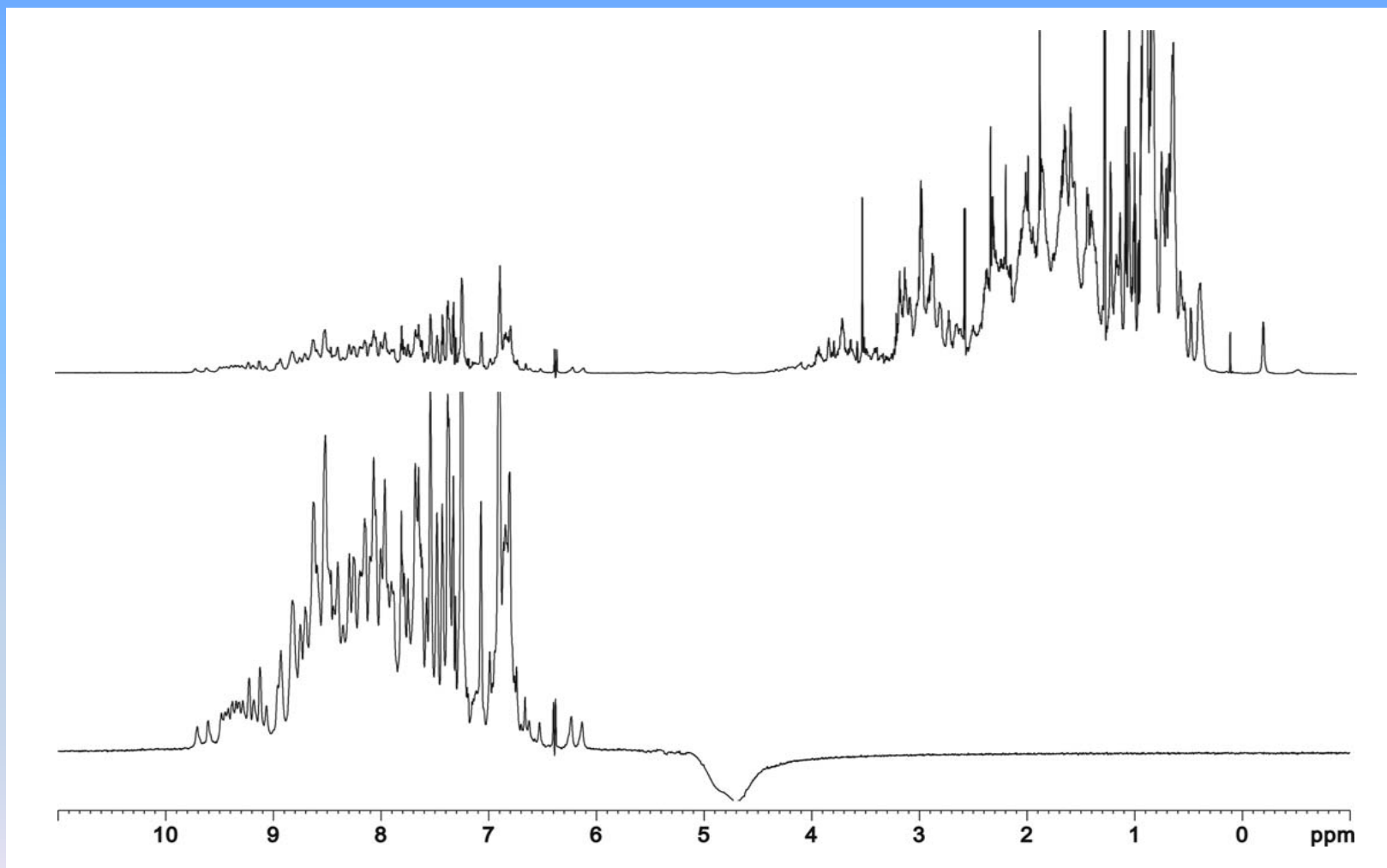
PC-9 2000 usec

rSNOB 667 usec

Test: 1D WATERGATE



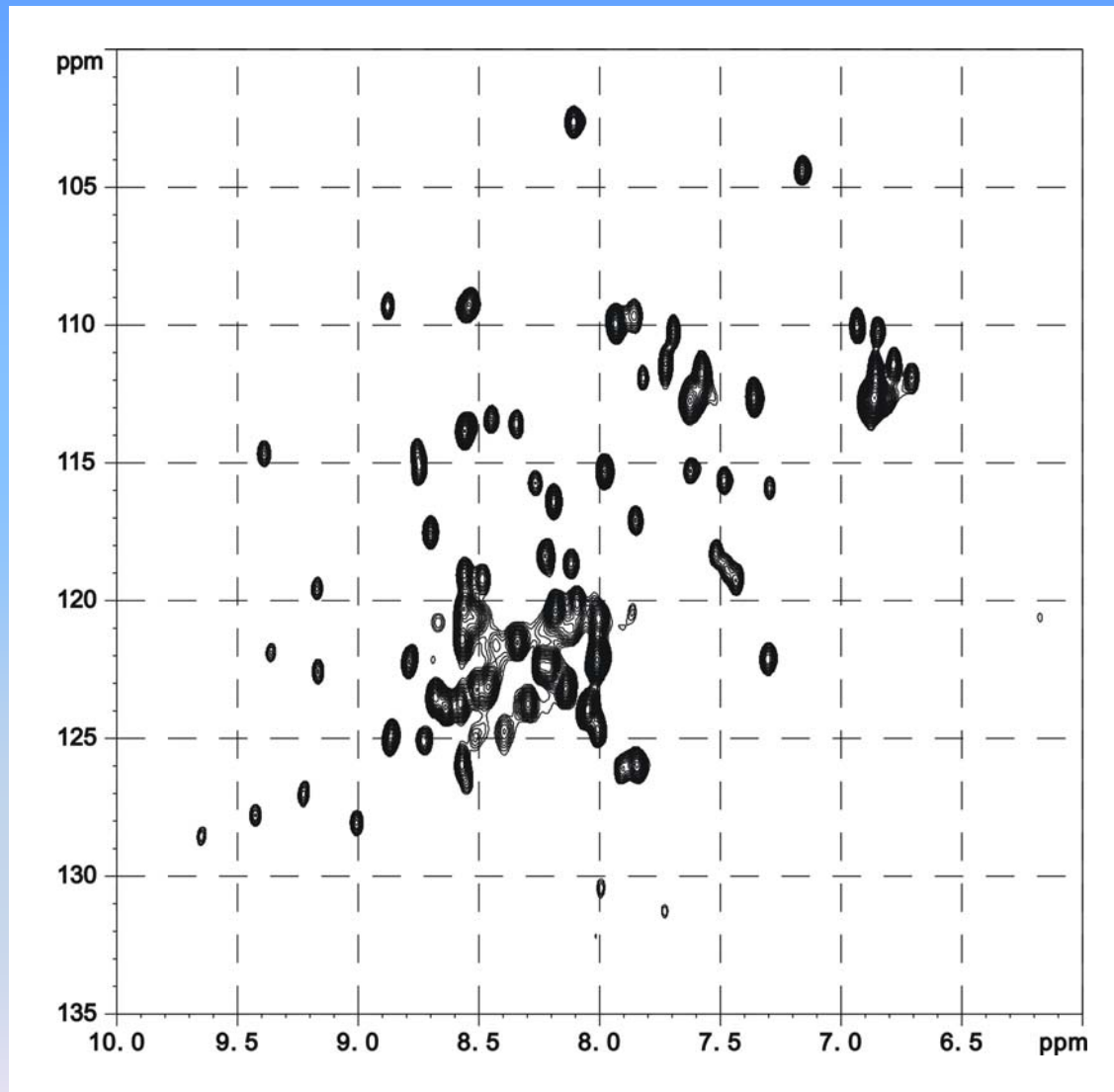
1D with a „SOFAST-WATERGATE“



Potential problems are probe heating because of the high repetition rate and the fact that the lock does not really have time to stabilize the field.

The probe heating might be especially problematic when using cryoprobes.

Bruker recommends to shorten the relaxation delay until the heating of the cryoprobe is getting close to zero



SOFAST-HMQC

of ^{15}N -Ubiquitin

900 MHz

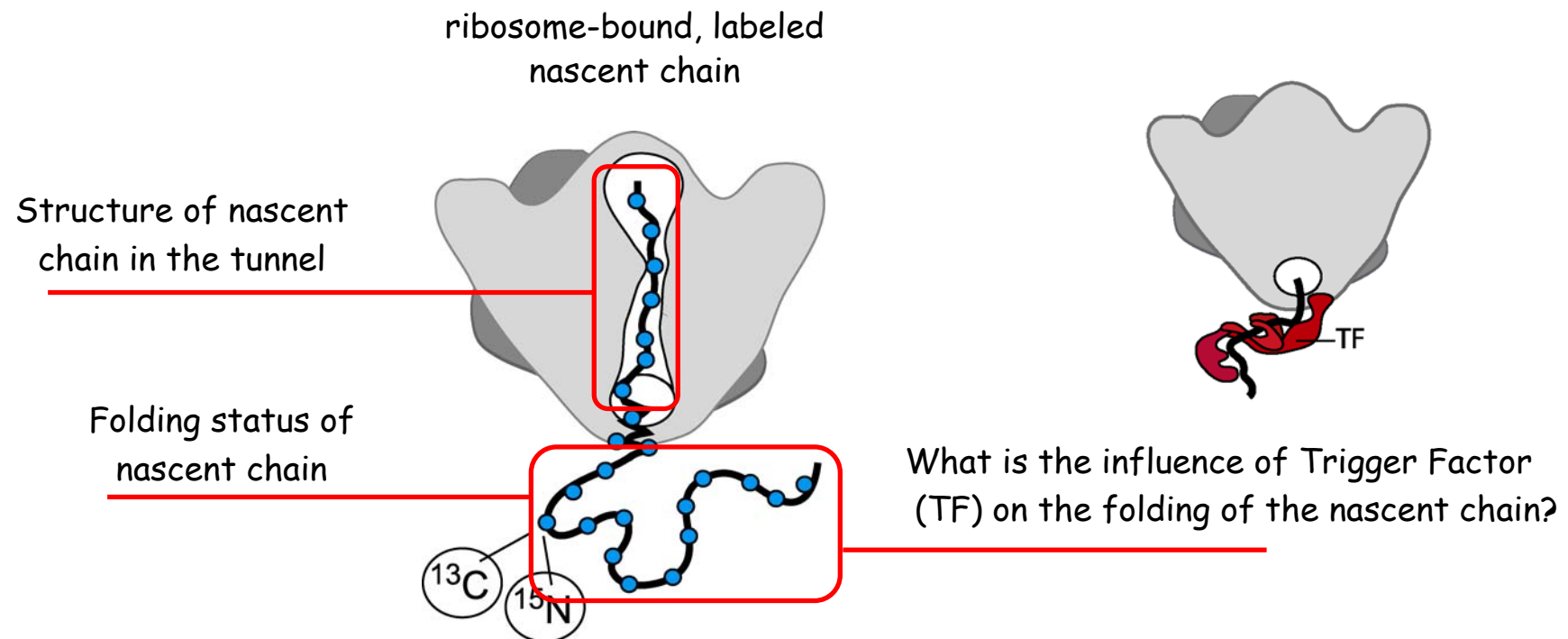
cryo-probe

2 scans, 128 FIDs

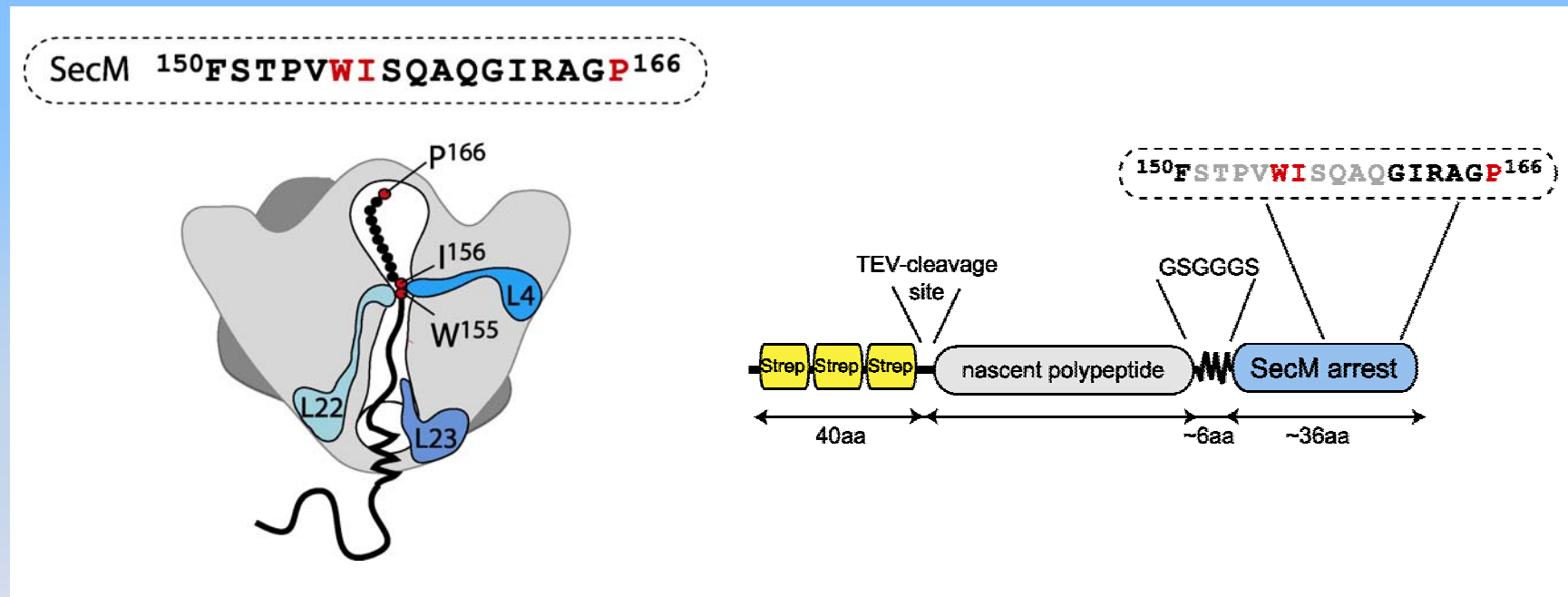
T=97 msec

Messzeit 25 sec

An alternative to shortening the measurement time is to improve the S/N in a given time for very demanding samples: the nascent chain bound to the ribosome

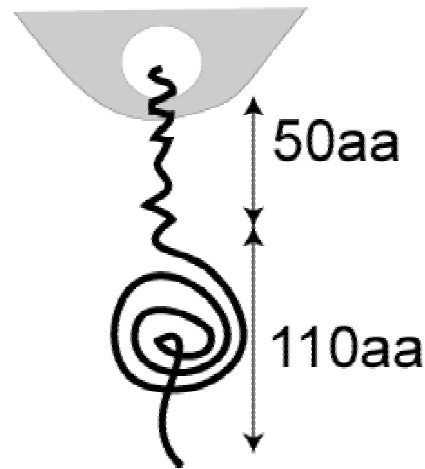


Samples have been prepared and the preparation optimized in the group of B. Buckau (Heidelberg)



We tried it with Barnase, that is known to give a folded protein when expressed alone

Barnase Δ 50



Things to test:

do we get signal ?

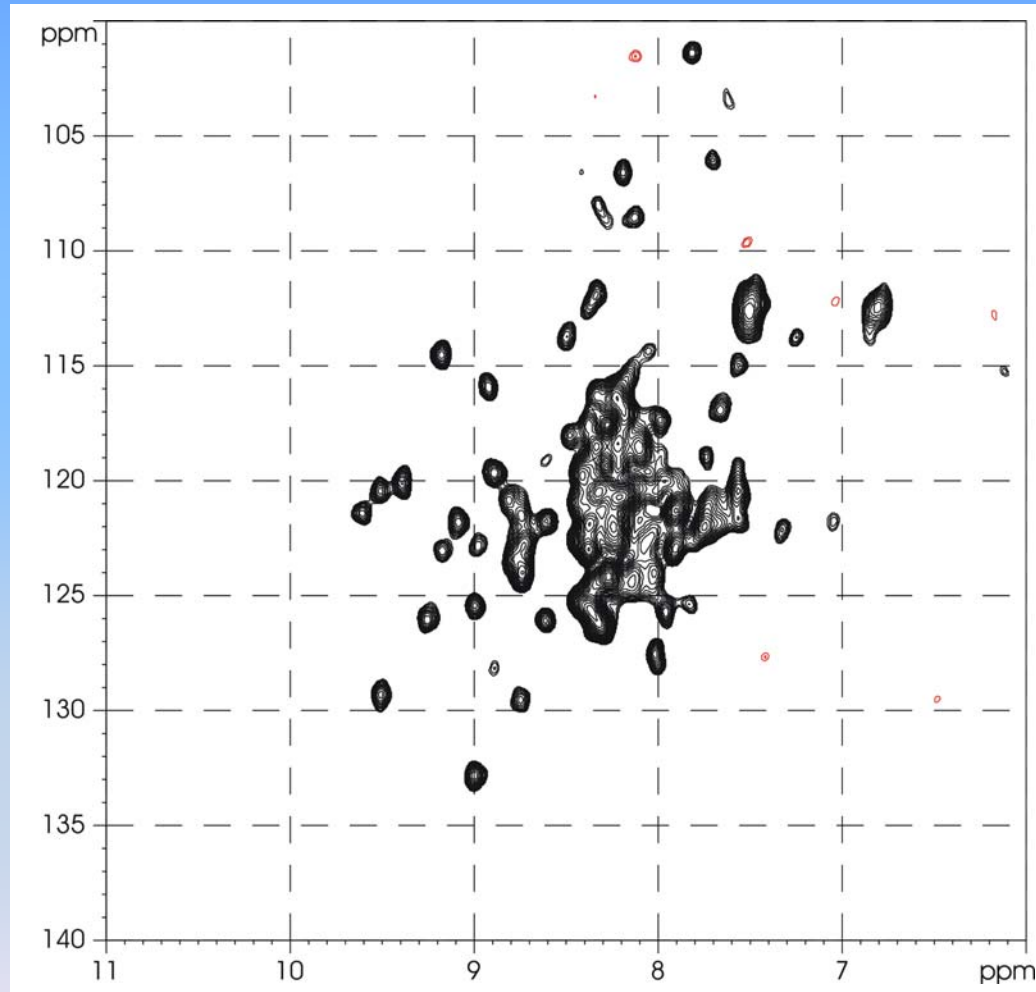
is the ribosome labeled as well ?

is barnase folded properly ?

how does it look when released

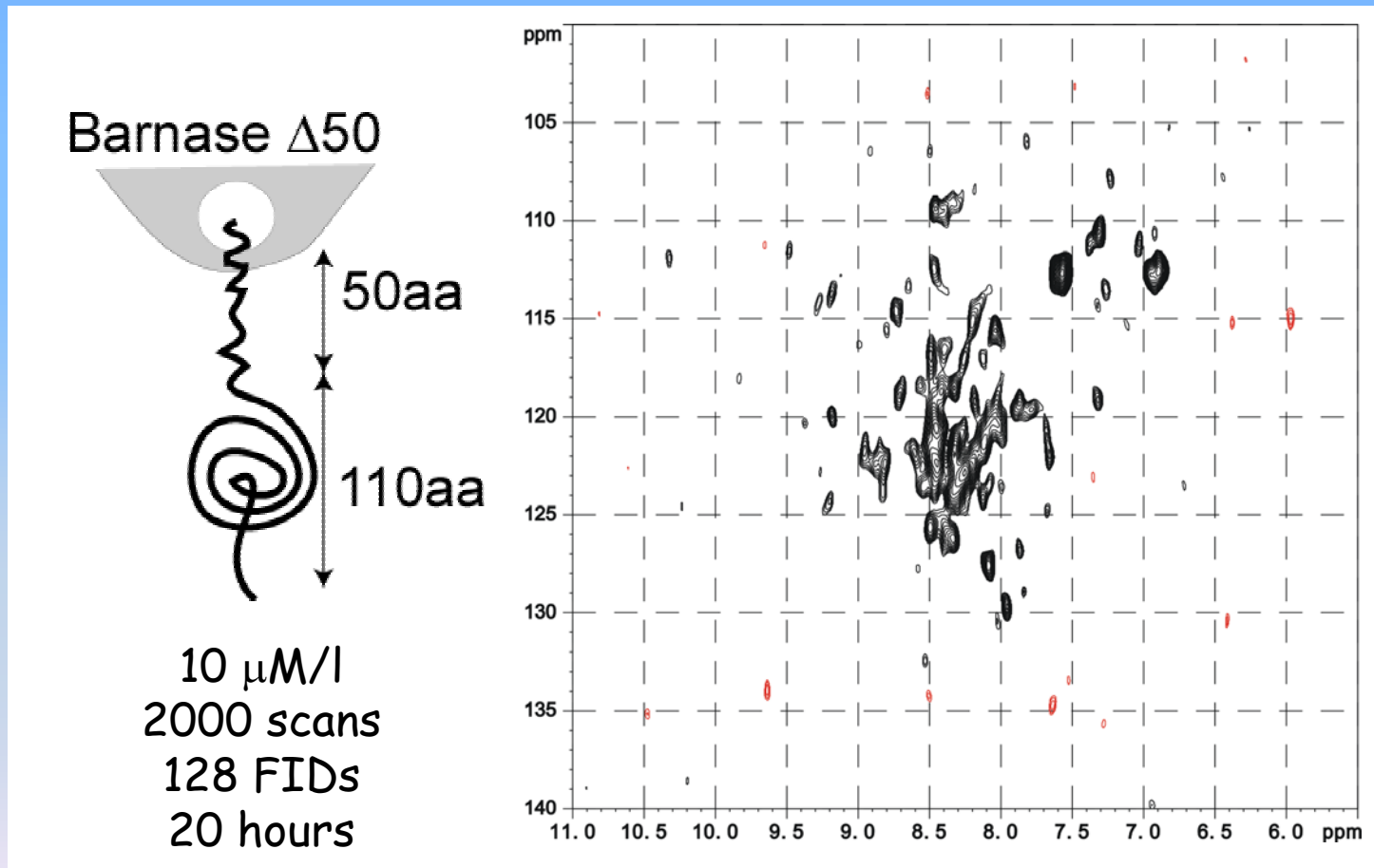
from the ribosome ?

First spectra were promissing, but....

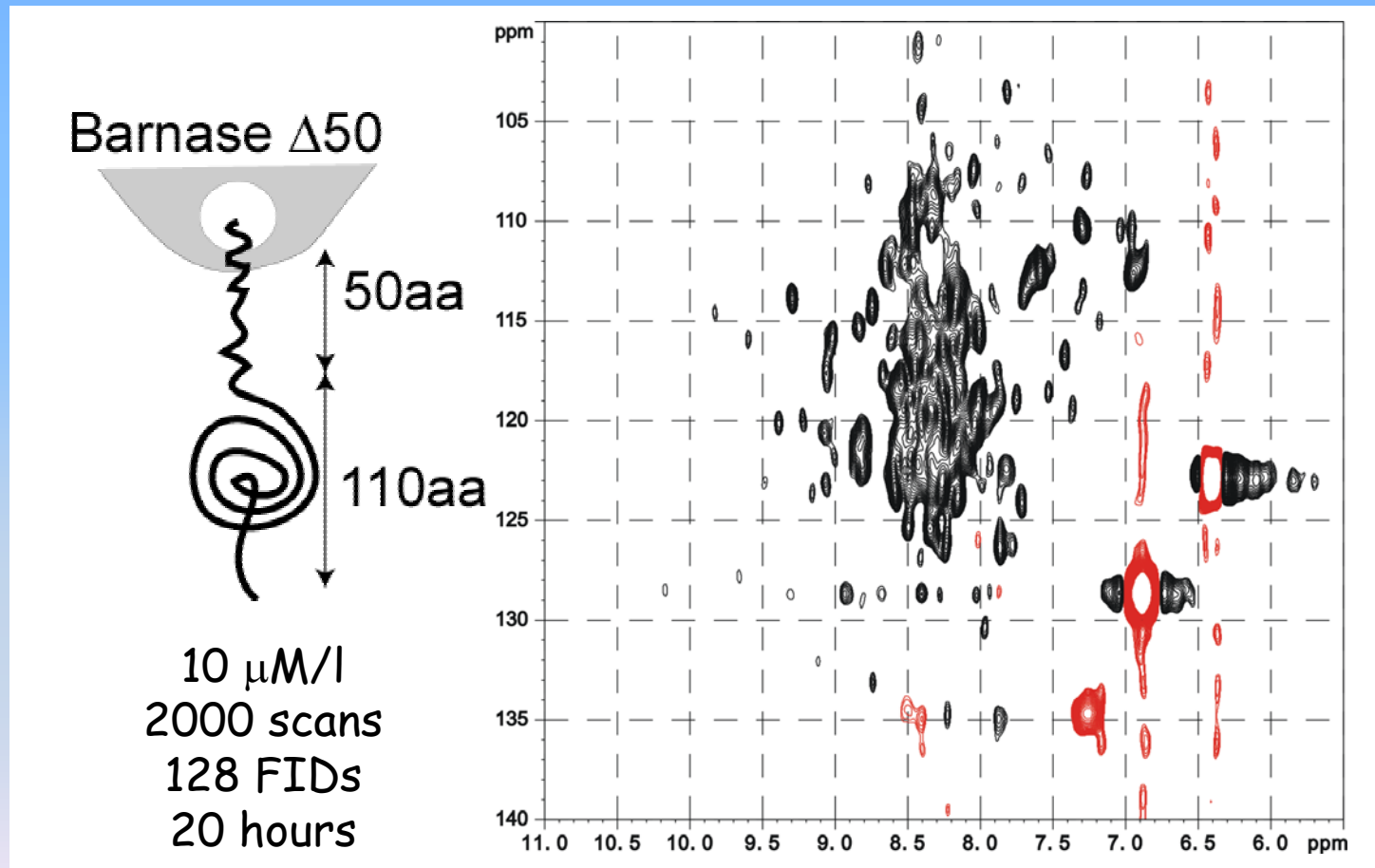


... turned out to be the ribosomal proteins L7/L27

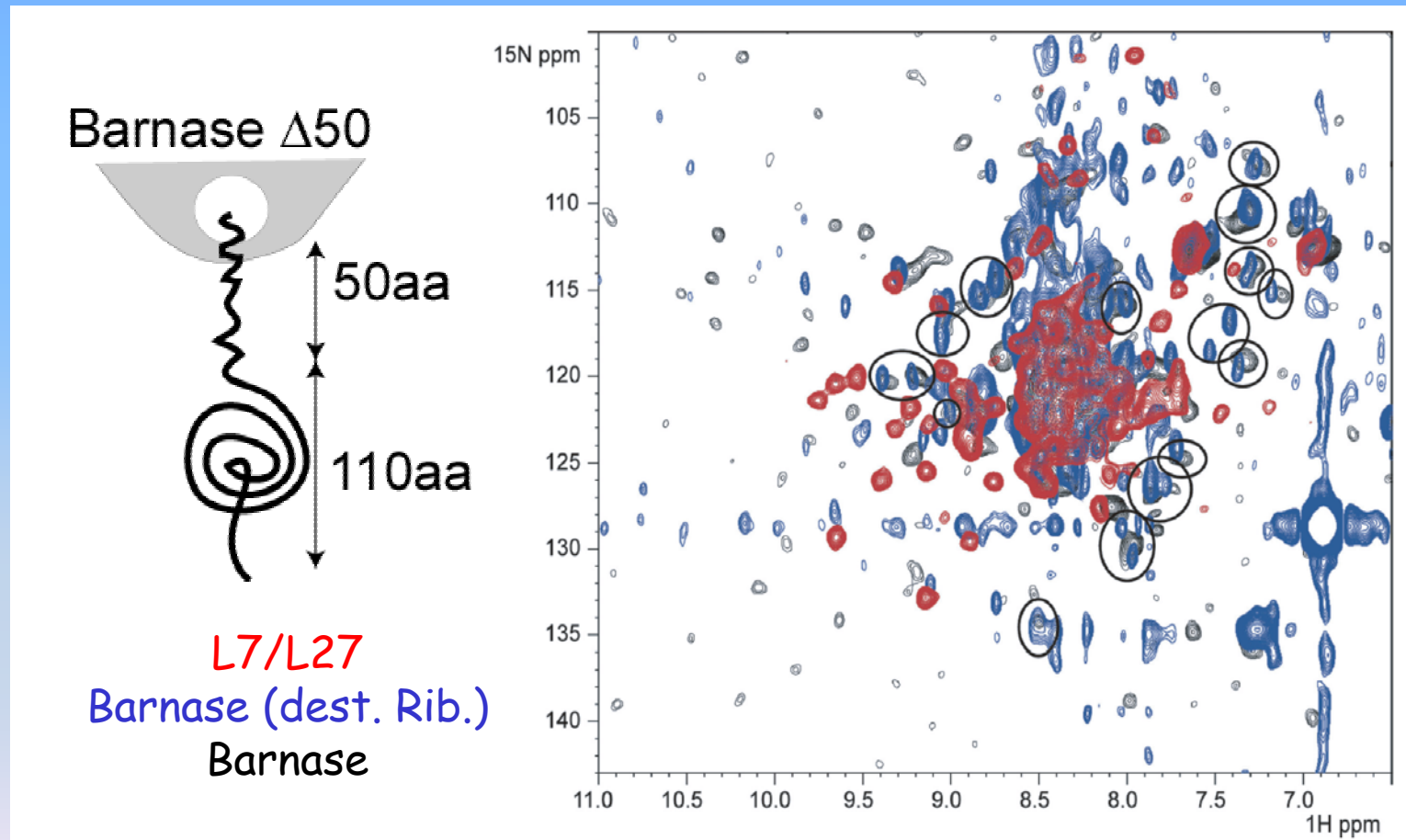
After optimization spectra were more
difficult to record, but it worked



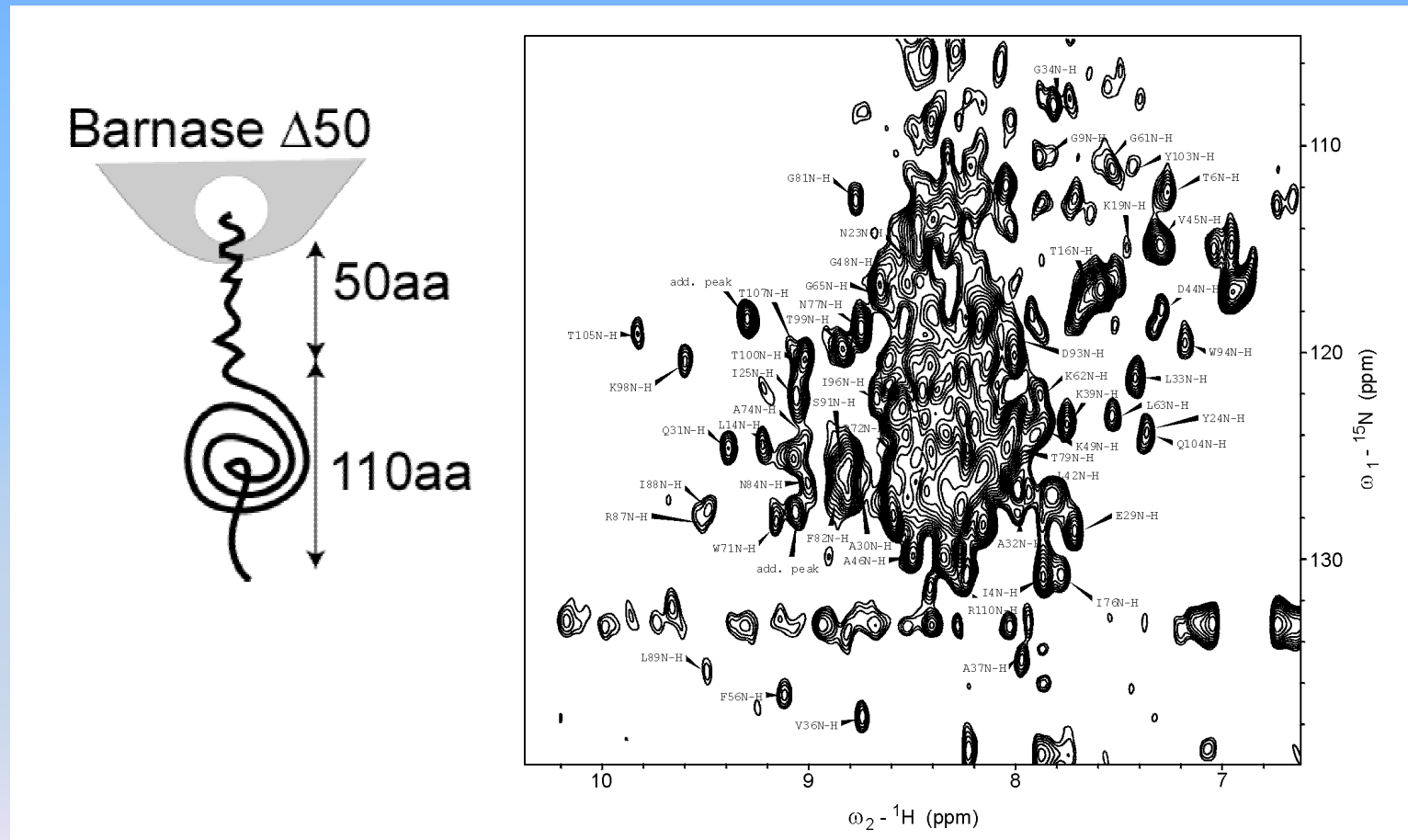
spectra were the ribosome is destroyed



The overlay looks good and with a look in the BMRB...



... one can do some assignments



acknowledgement

FMP

Monika Beerbaum

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Rainer Kümmerle (pdf)